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Target-based virtual screening and molecular dynamics approach to identify potential antileishmanial agents through targeting UvrD-like helicase ATP-binding domain

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Abstract

Background: About 0.7-1.0 million people worldwide have been suffering from Leishmaniasis. It falls under a neglected tropical disease (NTD) and is transmitted by biting infected female phlebotomine sandflies. The implication of “the NTD road map: together towards 2030” in the infection-prone regions worldwide has curtailed morbidity to a greater extent. However, limited options in antileishmanial oral and topical drugs must decipher more therapeutically efficacious agents to cure and eradicate the disease.

Methods: Virtual screening based on structure, docking, & molecular dynamics approaches were adopted to identify potential lead molecules against UvrD-like helicase of *Leishmania donovani* from the MCULE database. Lipinski rule of five, N/O atoms (1-15), number of rings (1-2), HBDs (4-5), and HBAs (5-10) were applied as initial filters of SBVS. AutoDock Vina and GROMACS packages were used for docking and MD simulations, respectively.

Results: Initial filters of SBVS workflow yielded 93885 ligand hits out of 100 plus million investigational ligands. Following the toxicology test, 28 ligands were gotten that were additional reduced to molecules (17) when accepted done the BOILED Egg model of the ADME. Six molecules were shortlisted with zero violation compliance of drug-likeness further than Lipinski RO5 viz., Egan, Veber, Muegge, Ghose, & bioavailability score having ΔG (-6.7 to -7.4 kcalmol⁻¹) lesser than reference inhibitor miltefosine (-4.9 kcalmol⁻¹). The stability of MCULE-5754880195-0-2 was found to be greater than the known inhibitor and ligand molecules mentioned above.

Conclusion: MCULE-5754880195-0-2 has all therapeutic features by way of an admirable oral drug molecule & could be encouraging in Leishmaniasis prevention & treatment.

Introduction

Leishmaniasis, which is regarded as one of the most neglected tropical diseases, is thought to be caused by more than twenty different *Leishmania* species. Over ninety sandfly species are known to spread *Leishmania* parasites. About 0.7-1.0 million new cases occur annually worldwide, especially in socially and financially deprived countries, although the progression of infection towards developing leishmaniasis is slow [1]. Three types of leishmaniasis exist. viz., visceral, mucocutaneous & cutaneous. The first one, visceral leishmaniasis (VL), is the fatal form of the disease, categorized via asymmetrical attacks of expansion of the spleen & liver, weight loss, fever, & anemia. The World Health Organization (WHO) claims, 2020 statistics >90% of VL incidence were reported from Brazil, Somalia, Yemen, Sudan, India, China, Ethiopia, Eritrea, Kenya, & South Sudan [2]. The second one is CL (cutaneous leishmaniasis) the utmost public form of the disease that causes severe skin wounds and scars mostly on the uncovered parts of the body. America, Mediterranean, Middle East, & Central Asia contribute over 90% of CL incidences compared to other countries [3]. The third one, mucocutaneous leishmaniasis (ML), is a comparatively mild form of the disease that destroys the membranes (mucous) of the nose, mouth, & throat. Maximum cases of ML arise in Brazil, Bolivia, Peru, & Ethiopia [4]. *Leishmania* parasites are transmitted when female phlebotomine sandflies infected suck the blood of the reservoir host to lay eggs. Several characteristics, viz., ecology of transmission sites, vector species, and present & past contact of the human populace to the parasite, influence the epidemiology of leishmaniasis. Besides humans, > 65 species (animal) have been reported as normal reservoir hosts of the leishmaniasis-causing organism [5]. People suffering from both leishmaniasis, and HIV are more prone to full blown medical disease & extraordinary revert & mortality rates, often seen in Ethiopia, Brazil, & India (Bihar) [6]. Moreover, socioeconomic conditions, environmental changes, population mobility, malnutrition, & climate change are the key risk factors associated with leishmaniasis. Prevention and control of leishmaniasis require holistic strategies because spreading the disease occurs through the complex biological system of the host revisor human, causing agents and vectors. Albeit persisting complexities in treatment options, primary diagnosis & effective quick treatment, effective disease observation, regulator of animal reservoir hosts, vector control, community mobilization, & consolidation partnerships are a few critical strategies for prevention and control of leishmaniasis. Moreover, the WHO's subsidized medicine price and donation program have accelerated disease prevalence reduction, curtailing the

associated disability and mortality to a certain extent [7].

According to classical concepts, the pathogenesis of leishmaniasis arises due to the inequity of T helper type 1 & T helper type 2 (TH1 & 2) cells [8]. Many complexities persist with the diagnosis of Leishmaniasis. Two approaches, direct (parasitic) and indirect (immunological), are standard for diagnosis. However, confirmatory inferences are not so easy in impoverished populations. Despite various problems, clinicians diagnose in the best way based on the history, epidemiology, clinical symptoms, and physical appearance of scars and lesions. There is limited oral therapeutics for the treatment of Leishmaniasis. Miltefosine and a few azoles viz., Fluconazole and Ketoconazole are used in used in systemic therapy. Miltefosine (DB09031) is the sole recognized oral drug to treat VL, CL, and ML. However, it was developed in 1980 as an anticancer agent, but now it has been repurposed as a broad-spectrum antileishmanial, antimicrobial, & phospholipid drug. Patients above the age of 12 are given it orally or topically. CDC (Centres for Disease Control and Prevention) has also suggested this one as a first line treatment for FLA (free-living amebae) infections, viz., Naeglerias is (principal amebic meningoencephalitis) & GAE (granulomatous amebic encephalitis) exceptional, fatal infection of the central nervous system (CNS) [9,10]. Drugs work through various molecular interactions within and between the cellular and signaling pathways which perform varied functions via finely tuned mechanism(s). Biological systems' molecular & cellular procedures continue in equilibrium concerning necessities and what not to do. Occasionally, several intrinsic and extrinsic factors hamper the equilibrium through either down-regulating or up-regulating the associated molecular targets, thus facilitating the onset of abnormalities [11,12,13]. Targeting enzymes of such cascaded pathways is a lucrative strategy to break the cycle of zoonotic pathogens. In the proposed work, we aim to identify investigational ligand(s) akin to miltefosine, which can inhibit the UvrD-like helicase (it is abbreviated as ULH hereafter), a putative protein of *Leishmania donovani* (strain BPK282A1) (EC: 3.6.4.12) (996 amino acids). The absence of ULH in humans makes it a promising therapeutic target. ULH consists of two domains: UvrD-like helicase ATP-binding (1-314 amino acids) and UvrD-like helicase C-terminal (309-678 amino acids). The inaccessibility of ULH's 3D structure in the PDB (protein data bank) prompts us to predict its structure using the Swiss-Model server of Expert Protein Analysis System (ExPASy) bioinformatics source portal (<https://swissmodel.expasy.org/>) [13]. The predicted model was assessed using the Ramachandran plot and

Qualitative Model Energy Analysis (QMEAN) tools [14]. Structure-based virtual screening by necessary search limits, including Pfizer's Lipinski rule of five (HBA 10; LogP 5; HBD 5; RO5: MW 500 Da) N/O atoms (1-15), and aromatic rings (1-2), was screen potential ligand (small molecule) from MCULE's massive digital tentative ligand library. The database (MCULE: <https://mcule.com/>) is a digital accessible drug innovation source portal with over a million synthesized accessible and purchased compounds for cell-based assays, preclinical, clinical trials. The SBVS-hits were docked through predicted ULH protein using AutoDock Vina (ADV), which is incorporated into the MCULE drug discovery platform, and then toxicophores were assessed. The Brain or Intestinal Estimate D (BOILED)-Egg model was used to test the HIA (Human Intestinal Absorption) & BBB (Blood Brain Barrier) permeability founded on two physicochemical parameters, namely TPSA (Topological Polar Surface Area) & WLOGP. The compounds that accepted the BOILED-Egg test were screened by drug-likeness attributes other than RO5, then through medicinal chemistry's PAINS (pan assay interference structure) & Brenk attentive exploration. The stability of top ligand hits was tested using the molecular dynamics (MD) modelling process and the aforementioned parameters [15]. To illuminate the binding capability of the found new prime molecule into the binding cavity of the therapeutic target protein ULH, a comparison analysis was performed amid the reference drug miltefosine & the finest prime molecule represented using the aforementioned methodologies.

Methods

Structure (3D) prediction & optimization of target protein

Swiss-Model web tool was used to 3D structure *Leishmania donovani* (strain BPK282A1). The model was built based on the template DNA helicase (PDB ID: 1PJR) of *Geobacillus stearothermophilus* (UniProt ID: P56255) [16]. Qualitative Model Energy Analysis (QMEAN) tool using consensus-based distance constraint, a.k.a. QMEANDisCo, and Ramachandran plot was used to evaluate the qualitative and stereochemical quality for the predicted model [17]. The target protein was modified by applying the CHARMM force field [18,19].

SBVS (Structure Based Virtual Screening)

MCULE, an online drug discovery platform, was employed for structure based virtual screening of experimental small ligands similar to miltefosine from its vast digital repository, which contains over 100 million synthetically accessible and purchasable molecules. In the structure based virtual screening

(SBVS) work procedure, the fundamental search limit complying with Pfizer's RO5, number of nitrogen and oxygen atoms, and number of aromatic rings was applied to screen chemical hits. Input values for the diversity range, sample size, & resemblance search threshold were 1000, 100, and 0.90, respectively. Small molecule fragments based robust 2D search algorithm, a.k.a. FP2 fingerprint of open babel, was assigned to execute the SBVS workflow [20,21].

Retrieval of structure (3D) & customization of reference drug

The structure (2D) of reference drug miltefosine (CID Number: 3599) in standard data format (SDF) was extracted from the database (NCBI & PubChem) [22, 23]. 2D to the 3D conversion of miltefosine was supported via the Biovia discovery studio visualizer (DSV). The ligand was actively reduced through a similar protocol by way of the target protein ULH [18, 24].

Molecular docking with AutoDock Vina

The AutoDock Vina (ADV) intrinsic toward the MCULE database (<https://mcule.com/>) was used for molecular docking concerning the target protein & Structure Based Virtual Screening (SBVS) - chemical hits. The PDB structure of the protein as an input file was delivered near the ADV edge of the portal (MCULE. Grid size in x (7.184 Å), y (31.4135 Å), and z (7.224 Å) directions were applied to shield the binding pockets of protein. The ADV settings (parameter) for binding mode per ligand & exhaustiveness were left at their default values. The free energy of binding (ΔG) was regarded as the most imperative factor for determining the optimal posture of ligands docked into the binding cavity of ULH [11, 25-27].

Toxicity investigation of virtually-screened hits

The existence of toxic moieties, fragments, & substructures in effectively open out ligand molecules unwelcomed in the human & eco-friendly environments was examined over and done with the Toxicity Checker tool of the MCULE database grounded on the difficult & robust SMARTS (SMILES (simplified molecular-input line-entry system) arbitrary target specification) algorithm [20].

Egan's BOILED-Egg filtration

The BOILED (Brain or Intestinal Estimate D) Egg model of the SwissADME implement was working to predict the BBB & HIA permeation of identified hits. The BOILED Egg relies on the 2 physicochemical descriptors, viz., WLOGP (≤ 5.88 as a reference value) & TPSA (≤ 131.6 as a reference value) for lipophilicity & seeming polarity, correspondingly & distinctive illustrative explanation of exactly how faraway a

molecule is from the perfect one for ideal preoccupation [28- 31].

Assessment of medicinal chemistry attributes

The medicinal chemistry properties include frequent hitters, often known as promiscuous compounds, which were anticipated using the pan assay interference structure (PAINS) observant option of SwissADME tools developed by Eli Lilly. Ruth Brenk's alert evaluated unwanted substructures, dyes, and hazardous compounds [32, 33].

Molecular dynamics (MD) simulation

GROMACS 5.1.2 was used to compute the stabilities of docked complexes of the best ligand-ULH and the reference medication miltefosine-ULH at 300K at the molecular mechanics' level. The ligands were divided as of their corresponding docked complexes through gmx grep module. The topology and forcefield consideration (parameter) files for the selected ligand hits were predicted by the CHARMM general force field (CGenFF) programme. The topologies for ULH were created using GROMACS 5.1.2's pdb2gmx modules. The CGenFF tool was used to retrieve the structural coordinates of screened hits and the reference drug miltefosine [34, 35, 36].

By a margin of 10 Å, all ligand and reference drug guaranteed complexes were soaked in a dodecahedron box of molecules (water). The gmx editconf module was used for making boundary circumstances. Using addition chloride & sodium ions via the gmx genion module, to maintain neutrality, the charges on the bound complexes were neutralised, preserving the physiological concentration (conc.) of 0.15 M. The system was then reduced for 250000 stepladders using the sharpest pedigree algorithm. The system temperature (temp.) was elevated from 0-300 K in the course of their equilibration of 5ns interval at persistent NVT & NPT. Subsequently the equilibration phase, the particle mesh was allocated through Ewald method [37, 38]. Different modules of the GROMACS package viz., gmx rmsf, gmx rms, gmx sasa & gmx ΔGsol, & gmx Rg, were used to depict the stability of lead molecules in terms of RMSD (root square deviation), RMSF (root mean-square fluctuation), SASA (solvent accessible surface area) the free energy of solvation (ΔGsol), & radius of gyration (Rg) plots [34, 39].

Results

Structure (3D) prediction of UvrD-like helicase, virtual screening and toxicity check

Swiss-Model server built the model of UvrD-like helicase (E9B9V1) using template DNA helicase (PDB ID: 1PJR) of *Geobacillus stearothermophilus* (UniProt

ID: P56255) (16), which target-template sequence similarity was found to be 27.72%. Average Model Confidence in terms of QMEANDisCo global score was found to be 0.54 ± 0.05 . Ramachandran plot exhibited that 92.02 % of residues of the model were located in its core region. Predicted models with >90% residues in the core region are considered promising [40]. The local confidence of the predicted model and Ramachandran plot is illustrated in Figures 1A and 1B, respectively.

Method (SBVS) was employed to identify small molecules from the MCULE's digital source of new ligands (>100 million). Basic filters of RO5 (zero violation), number of N/O atoms (range: 1-15), and aromatic rings (limit: 1-2) yielded 93885 ligand hits, followed by toxicity assessment through the SMARTS algorithm of SwissADME. Twenty-eight ligands turned out as non-toxic drug nominees, & the remaining 93,857 ligands were disallowed during rigorous toxicity checking.

The BOILED-EGG filtering technique

For instance, the name implies, the Egan model consists of two regions, i.e., yellow & white, depicting the physicochemical spaces for considerable BBB (blood-brain barrier) permeation & gastrointestinal (GI) absorption, a.k.a. human intestinal absorption (HIA). Seventeen molecules (out of 28) showed significant BBB penetration and HIA permeation. The BOILED-Egg extrapolation of 28 ligands & reference molecules, i.e., miltefosine, is revealed in Figure 1C. The blue color & red color dots denote P-gp (+ve & P-gp -ve molecules, which means some ligands that are a substrate of P-glycoprotein have effluxed out in the course of BBB dispersion, & non-substrate ligands can penetrate the brain membrane [28, 42].

Drug-likeness other than Lipinski RO5

Drug-likeness models viz., Muegge (Bayer), Veber (GSK), Ghose (Amgen & Egan (Pharmacia) Abbott bioavailability score (BS) were used to identify pro-drug molecules [30, 31, 42, 43, 44, 45]. These models qualitatively estimate the drug-likeness for a molecule (small) to develop future potential oral prime molecules. Out of seventeen, only six ligands, MCULE-5855858205-0-1, MCULE-8153392673-0-37, MCULE-5754880195-0-2, MCULE-4495139953-0-1, MCULE-7191245305-0-1, and MCULE-4952526932-0-1 obeyed the rules mentioned above with zero violation, and eleven ligands breached the law. Miltefosine exhibited 2, 1, 1, & 2 violations of Ghose, Veber, Egan, & Muegge models, correspondingly. Moreover, all six ligands and miltefosine exhibited a similar Abbott BS of 0.55.

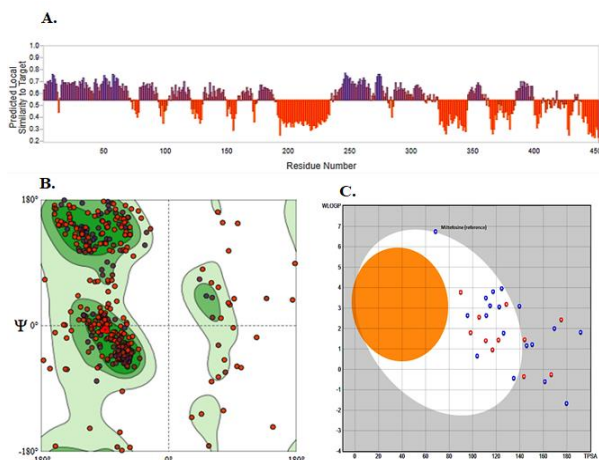


Figure 1: (A) Local quality estimate of the model through automated modelling of Swiss-model server. (B) Ramachandran plot of the predicted model of ULH showing 92.02% residues in the allowed region (favored/core region). (C) The BOILED-Egg assesses the passive GI absorption & BBB permeation of ligands & Miltefosine. None of the twenty-eight hits is located within the Egg yolk region, meaning no molecules exhibited BBB penetration. Eleven ligands are positioned outside the egg-white, meaning they do not show GI absorption. Miltefosine touches the egg-white line, meaning it is neither a brain penetrant nor a gastrointestinal absorber (negligible GI absorption). P-glycoprotein substrate and non-substrate were represented by dots in blue and red, respectively.

Docking simulation

Drug-likeness Using MCULE's ADV tool, successful ligands and Miltefosine were docked into the binding cleft of protein (target) ULH to analyze their binding affinities in terms of binding free energy (ΔG) ranging from -7.4 to -4.9 kcalmol⁻¹. Docked complexes of anticipated ligands & ULH were paralleled with reference molecule Miltefosine in terms of ΔG & type of connections. Miltefosine interacted into the binding pocket of ULH with a ΔG value of -4.9 kcalmol⁻¹ unveiling interactions with 20 residues via three different binding interactions viz., van der Waals (Vdw), carbon-hydrogen bond & Alkyl bonds (Figure 2A). Based on the ΔG criterion MCULE-5754880195-0-2 and MCULE-7191245305-0-1 were found as the top two ligands illustrating interactions with 13 residues through five (Vdw, HB, Alkyl, Pi-Alkyl, and Pi-Anion) and three (Vdw, HB, and Pi-Alkyl) binding interactions respectively (Figure 2B-C). Binding affinity and types of interaction involved in all six ligands passing through drug-likeness filtration are shown in Table 1.

S.No.	Ligands	ΔG (kcalmol ⁻¹)	UvrD-like helicase	
			Types of molecular interactions	
1.	MCULE-5855858205-0-1	-7.1	*Vdw, HB, CHB, Alkyl, Pi-Cation, and Pi-Pi T-shaped	
2.	MCULE-8153392673-0-37	-6.9	Vdw, Alkyl, Pi-Alkyl	
3.	MCULE-5754880195-0-2	-7.4	Vdw, HB, Alkyl, Pi-Alkyl, Pi-Anion	
4.	MCULE-4495139953-0-1	-6.7	Vdw, HB, Salt Bridge, Pi-Anion, and Alkyl	
5.	MCULE-7191245305-0-1	-7.3	Vdw, HB, and Pi-Alkyl	
6.	MCULE-4952526932-0-1	-6.9	Vdw, HB, Alkyl, Pi-Alkyl,	
7.	Miltefosine (reference drug)	-4.9	Vdw, CHB, and Alkyl	

*Van der Waals (Vdw), Predictable Hydrogen Bond (HB), Carbon Hydrogen Bond (CHB)

Table 1: Binding affinity of Drug-likeness succeeded ligands and type of interactions holding amino acid residues of UvrD-like helicase of *Leishmania donovani* (strain BPK282A1).

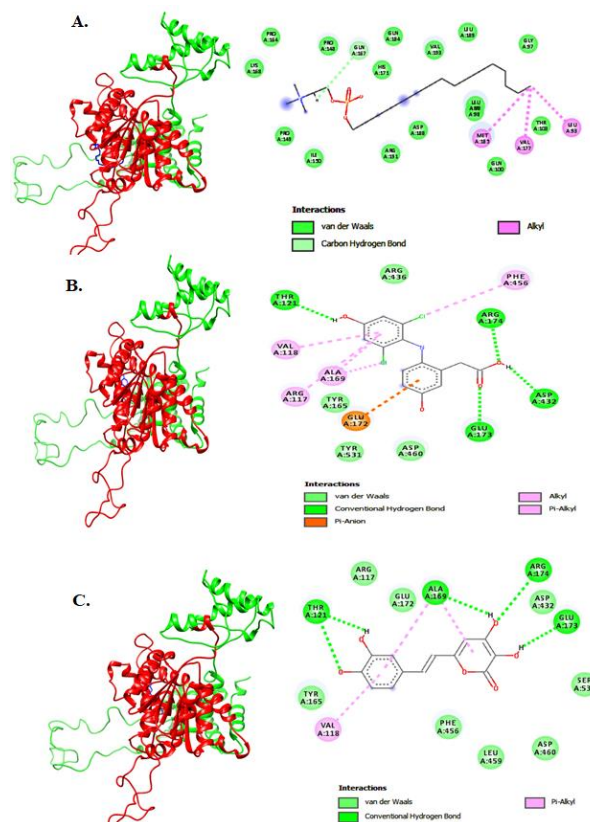


Figure 2: (A) ULH-Miltefosine complex. The figure (left) indicates the pose (3D) of the Miltefosine complex (blue sticks) docked to the binding pocket of ULH. The figure (right) illustrates the 2D pose of Miltefosine binding with dissimilar residues of ULH. ULH ATP-binding domain & ULH C-terminal are exhibited in red & green colors, correspondingly. (B) ULH- MCULE-5754880195-0-2 complex. The figure on the left depicts the 3D pose of the MCULE-5754880195-0-2 complex (blue sticks) docked to the binding pocket of ULH. The figure on the right depicts the 2D pose of MCULE-5754880195-0-2 binding by diverse residues of ULH. ULH ATP-binding domain & ULH C-terminal are exhibited in red & green colors, correspondingly. (C) ULH-MCULE-7191245305-0-1 complex. The figure on the left depicts the pose (3D) of the MCULE-7191245305-0-1 complex (blue sticks) docked to the binding pocket of ULH. The figure on the right depicts the 2D pose of MCULE-7191245305-0-1 binding with dissimilar residues of ULH. ULH ATP-binding domain & ULH C-terminal are exhibited in red & green colors, correspondingly.

Hydrogen bond formation and Stability investigation over MD simulations

The mention (reference) drug Miltefosine revealed nil conservative hydrogen bonds through binding interaction with the aim protein ULH, however MCULE-5754880195-0-2 and MCULE-7191245305-0-1 showed 4 and 5 hydrogen bonds respectively during interaction with ULH. These two ligands displayed a strong binding affinity by more negative ΔG values with target protein residues as compared to other selected ligands and inhibitor. As a result, only compounds with the greatest number of hydrogen bonds and the least amount of ΔG were chosen for MD investigations. [46, 47]. The stability of the top ligand hit docked complexes viz., MCULE-5754880195-0-2 & reference drug Miltefosine with ULH was investigated through MD simulations of 100 ns duration at Dell Workstation Precision 3440 using GROMACS (Groningen MACHine for Chemical Simulations) package. MD graphs for SASA, RMSF, RMSD, ΔG_{solv} , Rg, & HBs were plotted to assess the molecular docking stability of ligands & protein docked complexes [34].

RMSD (Root mean square deviation), RMSF (Root mean square fluctuation), SASA (Solvent accessible surface area) and Free solvation energy

The RMSD informs us about the stability of docked complexes. The mean RMSD for mention (reference) inhibitor Miltefosine (orange) & anticipated ligand hits MCULE-5754880195-0-2 (magenta) complexed with ULH was found 1.364 nm and 1.358 nm, correspondingly. The RMSD plot conceals that the stability of docked complex of ULH & MCULE-5754880195-0-2 is comparatively than inhibitor (Figure 3A).

The residues variations at numerous areas of the RMSF diagram (plot) are due to the binding interactions of Miltefosine & MCULE-5754880195-0-2 with ULH. Average residues fluctuation upon binding through Miltefosine (orange) and MCULE-5754880195-0-2 (magenta), was establish as 1.318 nm and 0.707 nm correspondingly (Figure 3B).

The SASA plot represents the surface area of the protein that is nearby to the solvent molecule. The usual (average) SASA values upon binding with Miltefosine (orange) & MCULE-5754880195-0-2 (magenta) was found as 94.452 nm² and 104.705 nm² respectively (Figure 3C).

The average ΔG_{solv} of ULH upon binding through Miltefosine (orange) & MCULE-5754880195-0-2

(magenta) was depicted as -126.761 kJ/mol/nm² & -144.476 kJ/mol/nm² correspondingly (Figure 3D).

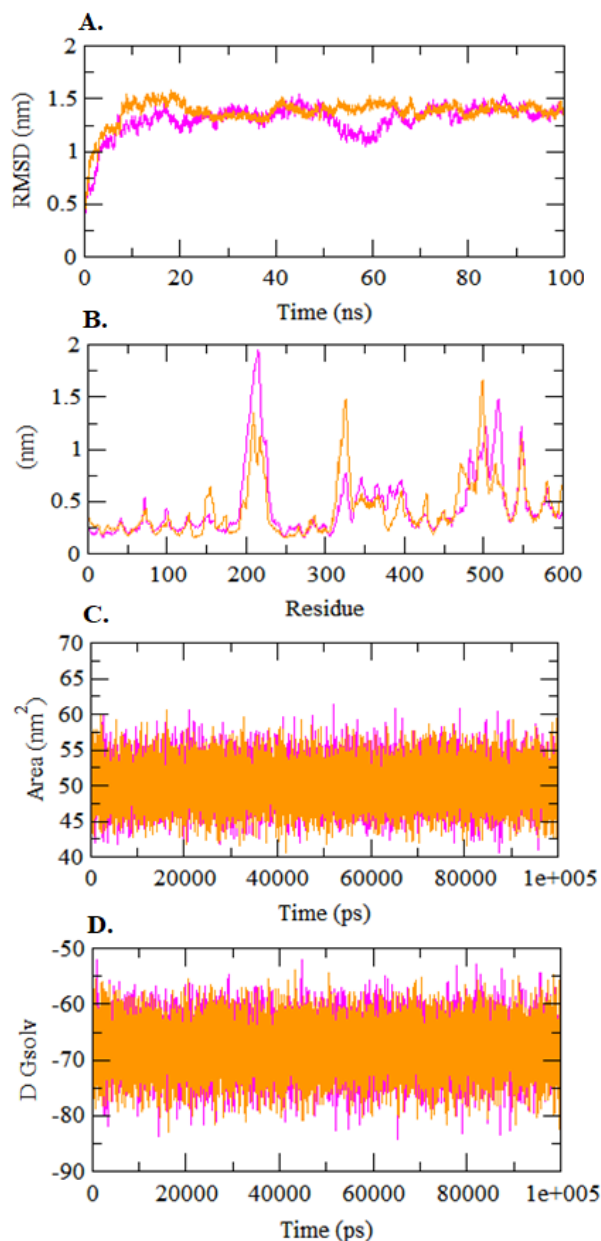


Figure 3: (A) Plotting the RMSD as a function of time. Orange & magenta signify values computed for ULH-Miltefosine & MCULE-5754880195-0-2 respectively. (B) RMSF diagram (plot) for ULH-Miltefosine (orange) and ULH- MCULE-5754880195-0-2 (magenta). (C) SASA plot for ULH-Miltefosine (orange) and ULH-MCULE-5754880195-0-2 (magenta). (D) ΔG_{solv} plot for ULH-Miltefosine (orange) and ULH- MCULE-5754880195-0-2 (magenta).

Gyration radius and HBs formation & deformation

The Rg masks the density of docked complexes and is inversely related to compactness. The ordinary (average) Rg values of docked complexes of Miltefosine (orange) & MCULE-5754880195-0-2 (magenta) were set

up as 6.016 nm and 5.971 nm correspondingly (Figure 4A).

The HB plot illustrates the number of hydrogen bond formations, deformation & stability during the all-inclusive process of simulations (MD).

Figure 4B indicates the HB plot for the docked complex Miltefosine-ULH and Figure 4C represents the nature of HB formation & deformation of ULH-MCULE-5754880195-0-2 during the all-inclusive course of molecular dynamics simulation of 100 ns period. Single HB is formed in ULH-Miltefosine, but they did not attain stability till the end of 50 ns MD simulation, though in the case of MCULE-5754880195-0-2, 5 HBs are being formed, that attains stability till the entire duration.

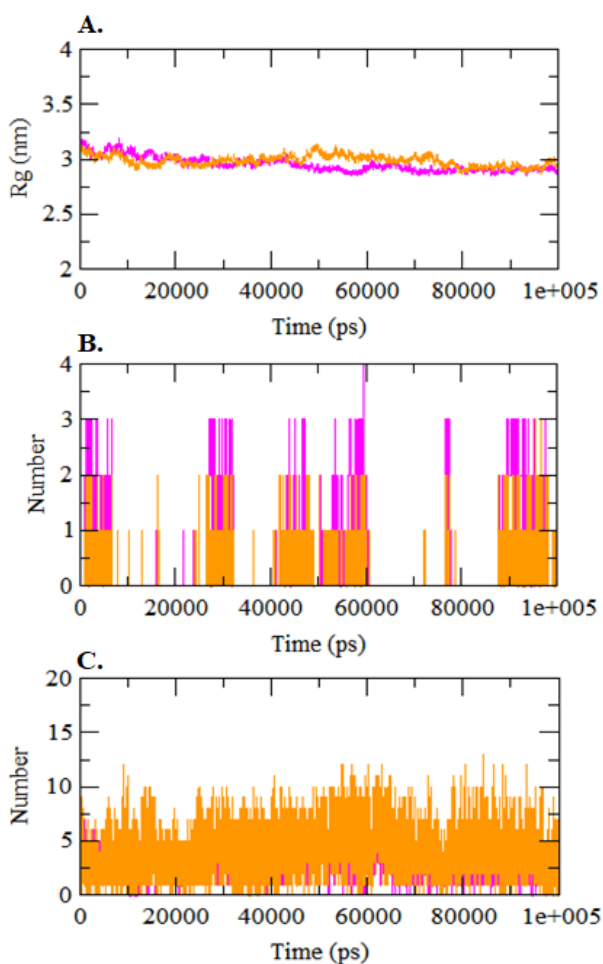


Figure 4: (A) Rg graphical (plot) for ULH-Miltefosine (orange) and ULH- MCULE-5754880195-0-2 (magenta). (B) HB plot illustrates the formation & deformation of H-bonds throughout interaction of Miltefosine by ULH. (C) HB plot indicates the formation & deformation of H-bonds for the duration of interaction of MCULE-5754880195-0-2 by ULH.

Discussion

The main goal of finding strong and targeted helicase inhibitors is to regulate how an organism can access its genetic material. Helicase inhibitors could theoretically be used to regulate any aspect of gene expression or replication. Nevertheless, the majority of current research focuses on developing helicase inhibitors that stop cancer cells or infectious infections from proliferating. Strong and targeted inhibitors of bacterial helicases, like the DnaB protein that acts at bacterial replication forks, or proteins involved in recombination, like RecBCD, may be used to create antibiotics. Cellular helicase inhibitors have the potential to function as antivirals, regulate cancer cells, or increase their susceptibility to chemotherapy [48,49,50]. The creation of medications to treat leishmaniasis is moving very slowly. Leishmaniasis can be treated with a limited number of medications, but managing the disease is made more difficult by toxicity, side effects, high costs, and the emergence of drug resistance. In order to prevent the spread of resistant *Leishmania* parasites and to help control leishmaniasis, it is necessary to develop new hits and leads [51]. The drug design and development field has seen a revolution in recent years thanks to nanotechnology. Compared to conventional chemotherapeutic options, it has demonstrated promise as a tool in parasitic diseases where the parasite relapses. Two ways nanotechnology has facilitated drug development are the creation of drug delivery systems and the nano formulation of medications that make them easy targets for macrophage phagocytosis, resulting in targeted drug delivery [52]. *Leishmania* adapts in advance to the environmental conditions of the host macrophage. Autophagy, which uses cathepsins and cysteine peptidases to regulate protein turnover, is an integral part of the life cycle of parasites and an essential step in their development and differentiation. This made these molecules viable therapeutic targets for future drug development [53].

ULH of *Leishmania donovani* is a putative therapeutic target against which drug design could be a lucrative strategy to prevent and treat Leishmaniasis. The predicted model of ULH depicts low sequence similarity with the selected template, but its more than ninety percent (90%) residues located in the most favored area (region) of the Ramachandran plot favor the useability in computer-aided drug design. High-throughput SBVS, profiling, physicochemical, toxicity properties, solubility, lipophilicity, drug-likeness, pharmacokinetics, medicinal chemistry attributes,

molecular docking, SASA, RMSF, RMSD, ΔG_{solv} , Rg & HBs evaluates determine that MCULE-5754880195-0-2 covers all therapeutic features that are superior to Miltefosine. In the purview of the above findings, MCULE-5754880195-0-2 may have emerged as an excellent oral drug candidate against Leishmaniasis. Experiments adopted are based on bioinformatics and computational tools having several possible research limits. Therefore, comprehensive wet-lab investigations are required to authenticate the in-silico findings of the research.

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Author Contributions

MMR, MZA and QA: designed and first drafted the manuscript. ZMS, JI and AA: data analysis, and revised the manuscript. MO and AFS: data collection and editing of the manuscript. BMA and NH: Final revision and editing of the manuscript. Authors read and approved the final version.

Conflict of Interest

The authors declare that there is no conflict of interest.

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