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Determination of total antioxidant content of methanolic extracts of *Cynara scolymus*, *Echinacea purpurea* and *Portulaca oleracea*

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Abstract

Background: The unique properties of medicinal plants particularly their effects on human health have attracted the attention of many researchers. The antioxidant properties of medicinal plants is a strong reason to use them in the food and pharmaceutical industries. Hence, the aim of this study was to evaluate the antioxidant properties of methanolic extracts of *Cynara scolymus*, *Echinacea purpurea* and *Portulaca oleracea*. Aerial parts of *C. scolymus*, *E. purpurea* and *P. oleracea* were dried and ground.

Methods: Then, plant samples were prepared using homogenizing plant powders in methanol solution. Finally, the total antioxidant capacity of the plants was assessed by ferric iron reducing antioxidant power (FRAP) assay.

Results: The results revealed that the total antioxidant capacity was found as 3.45, 1.16 and 1.68 mmol Fe²⁺/L for *C. scolymus*, *E. purpurea* and *P. oleracea*, respectively. Based on our results, *C. scolymus*, *E. purpurea* and *P. oleracea* showed a potent antioxidant activity.

Conclusion: It is recommended that utilization of *C. scolymus*, *E. purpurea* and *P. oleracea* in food and pharmaceutical industries could possibly possess beneficial health effects.

Introduction

Antioxidants have attracted the attention of scientists for their numerous benefits [1, 2]. Lack of antioxidants can induce oxidative stress and develop various diseases including neoplasms, cardiovascular diseases, neurodegenerative diseases and inflammatory disorders [3-5]. Oxidative stress refers to conditions in which the production of free radicals particularly the generation of reactive oxygen species (ROS) overcomes their neutralization by antioxidants [6-8]. ROS are naturally produced in low amounts in the body, but their overproduction can induce mitochondrial dysfunction, endoplasmic reticulum stress and eventually damage macromolecules and tissue injury [5-9]. To solve this problem, the use of antioxidants, especially oral antioxidants with natural origin is a useful solution [10]. It has been reported that the dietary intake of natural plant antioxidants has an inverse relationship with the incidence of diseases [11,12]. Medicinal plants have exerted healing effects due to their ability to the production of a wide range of antioxidant compounds such as polyphenols, terpenoids and vitamins [13,14]. The use of medicinal plants to treat various diseases has a long history [15,16]. This has led to use medicinal plants and the plant derived antioxidant compounds in finding treatment strategies to successfully treatment of human diseases [17,18].

Cynara scolymus is one of the medicinal plants with high antioxidant properties which grow in Mediterranean area. It is a perennial plant belongs to Asteraceae family which commonly known as artichoke. In addition to its edible use as a vegetable, *C. scolymus* has a long history in the treatment of diseases. Antioxidant constituents including phenolic acids and flavonoids are abundant in *C. scolymus*. Apigenin and luteolin are the most important bioactive compound which found in artichoke. Numerous healing properties, both in traditional medicine and in new research, such as antimicrobial, anticancer, hypoglycemic and hepatoprotective properties have been listed for *C. scolymus* [19]. *Echinacea purpurea* is a perennial plant belonging to Asteraceae family. *E. purpurea* could exert several biological and pharmacological effects including antioxidant, antimicrobial, anti-inflammatory, anti-tumoral and immunomodulatory properties. Researchers have indicated that the presence of bioactive compounds such as caffeic acid, caftaric acid, cichoric acid, echinacoside, alkylamide, α -pinene and limonene are associated with the healing effects of *E. purpurea* [20]. *Portulaca oleracea* is an annual plant belongs to the Portulacaceae family. It could grow in the tropical and subtropical regions including United States and Mediterranean countries. *P. oleracea* is used as a folk remedy via exerting antioxidant, antibacterial and anti-inflammatory effects. Flavonoids, alkaloids and

terpenoids are the most well-known bioactive compound extracted from *P. oleracea* [20]. Hence, the aim of this study was to evaluate the antioxidant properties of *C. scolymus*, *E. purpurea* and *P. oleracea*.

Methods

Collection of Plant Materials

The aerial parts of *C. scolymus*, *E. purpurea* and *P. oleracea* were collected from Dehloran county of Ilam Province, Southwest of Iran, in March 2022. The plants were identified according to the morphological features of Ilam Province Plant Flora at the Biotechnology and Medicinal Plants Research Center, Ilam University of Medical Sciences, Ilam, Iran. Collected plants were air dried in the shade and then were ground and used for antioxidant evaluation.

Plant Sample Preparation

After drying the plant, 1 g of the dry powder of the studied plants was homogenized using 100 ml of methanol solution and was shaken in the same solution for 6 hours. The resulting solution was then poured into a plastic falcon and centrifuged at 6000 rpm for 10 minutes. The resulting solution was used as a sample [21].

Determination of Antioxidant Activity

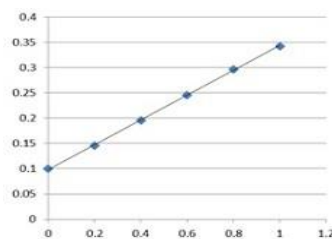
The total antioxidant capacity of the plants was assessed by ferric iron reducing antioxidant power (FRAP) assay [21].

Stock Solution Preparation

2.2 mL of R2b solution was added to the parent bottle R2a and vortexed until complete dissolution and R2 solution was obtained. Then, the R2 solution was mixed in a ratio of 1: 1 and after vortexing, 5 times its volume was added to R1 solution. The resulting solution is the stock solution of an antioxidant kit [21].

Standard Solution Preparation

Standard solution at 0, 0.2, 0.4, 0.6, 0.8 and 1 was also prepared. The linear equation obtained from the different concentrations of the standard solution is illustrated in Figure 1.



$$Y = 0.2447X + 0.0988$$

$$R^2 = 0.9997$$

Figure 1: Linear equation obtained from different concentrations of standard solution.

Procedure

First, 5 µL of the prepared plant solution was added to each well and then 250 µL of the prepared working solution was added to each well containing the plant solution. The microplate was then incubated at 35-50 °C for 30 minutes and finally read at 570 nm with the ELISA reader [22].

Results

As shown in Table-1, the results revealed that the total antioxidant capacity was found as 3.45, 1.16 and 1.68 mmol Fe²⁺/L for *Cynara scolymus*, *Echinacea purpurea* and *Portulaca oleracea*, respectively.

Plant	Total Antioxidant Capacity
<i>Cynara scolymus</i>	3.45 mmol Fe ²⁺ /L
<i>Portulaca oleracea</i>	1.68 mmol Fe ²⁺ /L
<i>Echinacea purpurea</i>	1.16 mmol Fe ²⁺ /L

Table 1: Total antioxidant capacity of *Cynara scolymus*, *Echinacea purpurea* and *Portulaca oleracea*.

Discussion

Oxidation reactions not only are a harmful process in the human body but also for many food products. Excess in generation of ROS is called oxidative stress which could induce development of various damaging conditions such as incidence of cancer, diabetes, inflammatory diseases and coronary heart diseases[23]. Meanwhile, antioxidants can protect the body against damage caused by oxidative stress as a shield. Antioxidants play their protective role by neutralizing free radicals by donating electrons to them and stopping the auto-oxidative chain reaction caused by ROS [24]. One way to supply exogenous antioxidants is by consuming rich sources of antioxidants. Medicinal plants have always been considered by researchers due to having rich sources of bioactive compounds with abundant biological and pharmacological properties particularly antioxidant effect[25]. Therefore, the present study was designed to evaluate total antioxidant capacity of *C. scolymus*, *E. purpurea* and *P. oleracea* using FRAP assay.

The results of our study indicated that the total antioxidant activity of *C. scolymus* was obtained as 3.45 Fe²⁺/L. The presence of bioactive constituents such as phenolic compounds especially phenolic acids and flavonoids play role in increasing total antioxidant activity of medicinal plants[26]. *C. scolymus* is recognized as a plant containing potent antioxidants. *C. scolymus* has a high rank in terms of containing antioxidant compounds among different medicinal plants.

In this regard, M. Ben Salem and the colleagues reported considerable antioxidant potential of *C. scolymus*. The results of their study showed that the value of total antioxidant activity of ethanolic extract of *C. scolymus* leaves using FRAP assay was obtained as

527.79 mol Fe (II)/g DW. They concluded that *C. scolymus* possessed significant antioxidant capacity. Although there was a difference in the values obtained, two studies confirmed the antioxidant effects of this plant. There were differences due to the differences in the geographical area, plant growth and ecological conditions and type of plant sample preparation [27].

In another study, Emanuel and the colleagues confirmed the ability of *C. scolymus* to scavenge free radicals. They showed that the presence of high quantities of phenols and flavonoids are related to capable antioxidant activity of *C. scolymus* [28]. In our study, the findings demonstrated that the total antioxidant capacity of *E. purpurea* was found as 1.16 Fe²⁺/L. *E. purpurea* exerts a remarkable antioxidant effect due to the presence of bioactive compounds including cichoric acid.

In a study, K.O. Mohamed Sharif and the colleagues investigated biological effects of extracts of *E. purpurea*. They found that the aqueous extract of *E. purpurea* leaves exhibited strong antioxidant activity. In this study, the value of total antioxidant activity of aqueous extract of *E. purpurea* leaves using FRAP assay was found as 196 mg TE/g which represents a high antioxidant capacity [29].

In another study, A. Wojdyło and the colleagues investigated antioxidant activity of a series of selected herbs including *E. purpurea*. In their study, the value of total antioxidant activity of aqueous extract of *E. purpurea* leaves using FRAP assay was found as 94.6 µmol Trolox/100 g dw [30]. The results of another part of our study indicated that the total antioxidant capacity of *P. oleracea* was found as 1.68 Fe²⁺/L. *P. oleracea* is known by its high nutritive value and healing properties. The antioxidant effect of *P. oleracea* could be related to the high total phenolic and flavonoid content.

Free radicals are toxic products of oxygen metabolism that contain at least one unpaired electron. Excessive production of free radicals during a process called oxidative stress causes damage to different parts of the cell such as proteins, DNA and cell membrane [31-37]. These irreversible damages lead to changes in cell function or cell death. Oxidative damage to vital molecules ultimately leads to chronic diseases such as heart disease, cancer, diabetes, Alzheimer's, Parkinson's, arthritis and infertility [38-43].

The human body needs both types of oxidants (free radicals) and antioxidants for normal metabolism, signal transmission, and regulation of cellular activities [44-45]. Medicinal plants and herbal antioxidants can contain compounds that can neutralize many diseases [46-62].

In a similar study, Md. Kamal Uddin and the colleagues evaluated the antioxidant properties of *Portulaca*

oleracea. They showed that the value of total antioxidant activity of extract of *Portulaca oleracea* using FRAP assay was found as 4.3 mg GAE/g. They concluded that this antioxidant property could relate to its high phenolic and flavonoid content [63]. In another study, it has been shown that *Portulaca oleracea* particularly its ornamental type could exert considerable antioxidant property. They revealed that the value of total antioxidant activity of extract of *Portulaca oleracea* using FRAP assay ranged from 7.39±0.08 to 104.2±6.34 μ mol TE/gDW [64]. Taking together, we concluded that the three chosen medicinal plants including *C. scolymus*, *E. purpurea* and *P. oleracea* shows a potent antioxidant property and this could be a reason for their use in the food and pharmaceutical industries. Based on our findings, it is recommended that utilization of *C. scolymus*, *E. purpurea* and *P. oleracea* could possibly possess beneficial health effects.

Competing Interest

The authors declare that there is no conflict of interest.

Author Contributions

Samira Shokri: Data analysis

Kourosh Saki: Monitoring of research

Yeganeh mazaheri and Samira Shokri: Drafting

Gholamreza Jahed khaniki: Manuscript scanning

Samira Shokri: Data evaluation

References

- Hormozi M, Marzijerani AS, Baharvand P. Effects of Hydroxytyrosol on Expression of Apoptotic Genes and Activity of Antioxidant Enzymes in LS180 Cells. *Cancer Management and Research*, (2020);12:7913.
- Nori-Garavand R, Hormozi M, Narimani L, Beigi Boroujeni N, Rajabzadeh A, Zarei L, et al. Effect of selenium on expression of apoptosis-related genes in cryomedia of mice ovary after vitrification. *BioMed Research International*, (2020);3(2): 1.
- Ahmadvand H, Tavafi M, Khosrowbeygi A, Shahsavari G, Hormozi M, Beyranvand K, et al. Amelioration of lipid peroxidation in vivo and in vitro by Satureja khozestanica essential oil in alloxan-induced diabetic rats. *Journal of Diabetes & Metabolic Disorders*, (2014);13(1):1-6.
- Hasanvand A, Pirzadroozbahani N, Ahmadizar F, Kharazmkia A, Mir S, Baharvand PA, et al. Evaluation of the antioxidant effects of zolpidem in the rat model of cisplatin-induced nephrotoxicity. *Journal of Renal Injury Prevention*, (2018);7(4):235-9.
- Mohammadrezaei Khorramabadi R, Anbari K, Salahshoor MR, Alasvand M, Assadollahi V, Gholami M. Quercetin postconditioning attenuates gastrocnemius muscle ischemia/reperfusion injury in rats. *Journal of Cellular Physiology*, (2020);235(12):9876-83.
- Bagheri S, Sarabi MM, Khosravi P, Khorramabadi RM, Veiskarami S, Ahmadvand H, et al. Effects of Pistacia atlantica on oxidative stress markers and antioxidant enzymes expression in diabetic rats. *Journal of the American College of Nutrition*, (2019);38(3):267-74.
- Bagheri S, Khorramabadi RM, Assadollahi V, Khosravi P, Cheraghi Venol A, Veiskarami S, et al. The effects of pomegranate peel extract on the gene expressions of antioxidant enzymes in a rat model of alloxan-induced diabetes. *Archives of Physiology and Biochemistry*, (2021):1-9.
- Bagheri S, Moradi Sarabi M, Gholami M, Assadollahi V, Mohammadrezaei Khorramabadi R, Hadipour Moradi F, et al. D-limonene in diabetic rats. *J Renal Inj Prev*, (2021);10(3):e24-e.
- Sarabi MM, Khorramabadi RM, Zare Z, Eftekhar E. Polyunsaturated fatty acids and DNA methylation in colorectal cancer. *World journal of clinical cases*, (2019);7(24):4172.
- Ahmadvand H, Babaeenezhad E, Nasri M, Jafaripour L, Khorramabadi RM. Glutathione ameliorates liver markers, oxidative stress and inflammatory indices in rats with renal ischemia reperfusion injury. *Journal of Renal Injury Prevention*, (2018);8(2):91-7.
- Zendedel A, Gharibi Z, Anbari K, Abbaszadeh A, Khorramabadi RM, Soleymaninejad M, et al. Selenium ameliorate peripheral nerve ischemic-reperfusion injury via decreased TNF- α . *Biological trace element research*, (2017);176(2):328-37.
- Hasanvand A, Kharazmkia A, Mir S, Khorramabadi RM, Darabi S. Ameliorative effect of ferulic acid on gentamicin-induced nephrotoxicity in a rat model; role of antioxidant effects. *Journal of Renal Injury Prevention*, (2018);7(2):73-7.
- Delfani S, Mohammadrezaei-Khorramabadi R, Abbaszadeh S, Naghdi N, Shahsavari S. Phytotherapy for Streptococcus pyogenes. *Journal of Pharmaceutical Sciences and Research*, (2017);9(5):513.
- Delfani S, Mohammadrezaei-Khorramabadi R, Ghamari S, Boroujeni RK, Khodabandeloo N, Khorzoughi MG, et al. Systematic review for phytotherapy in Streptococcus Mutans. *Journal of Pharmaceutical Sciences and Research*, (2017);9(5):552.
- Ghamari S, Mohammadrezaei-Khorramabadi R, Mardani M, Shahsavari S. An overview of the most important medicinal plants with anti-toothache property based on ethnobotanical sources in Iran. *Journal of Pharmaceutical Sciences and Research*, (2017);9(6):796.
- Fazeli-Nasab B, Shahraki-Mojahed L, Dahmardeh N. Evaluation of Antimicrobial Activity of Essential Oil and Ethanolic Extract of 10 Medicinal Plants on Rathayibacter tritici and Xanthomonas translucens. *Plant Biotechnol Persa*, (2022); 4 (1): 9-17.
- Beyranvand F, Gharzi A, Abbaszadeh A, Khorramabadi RM, Gholami M, Gharravi AM. Encapsulation of Satureja khuzistanica extract in alginate hydrogel accelerate wound healing in adult male rats. *Inflammation and regeneration*, (2019);39(1):1-12.
- Pourjabali M, Mohammadrezaei-Khorramabadi R, Abbaszadeh S, Naghdi N, Naji-Haddadi S, Bahmani F. Medicinal plants used for hypertension. *Journal of Pharmaceutical Sciences and Research*, (2017);9(5):537.
- Miraj S, Kiani S. Study of therapeutic effects of Cynara scolymus L.: A review. *Der Pharmacia Lettre*, (2016);8(9):168-73.
- Manayi A, Vazirian M, Saeidnia S. Echinacea purpurea: Pharmacology, phytochemistry and analysis methods. *Pharmacognosy reviews*, (2015);9(17):63.
- Dokhani N, Nazer MR, Shokri S, Darvishi M. Determination and Evaluating the Antioxidant Properties of Ziziphium nummularia (Burm.f.) Wight & Arn., Crataegus pontica K.Koch and Scrophularia striata Boiss. *Egyptian Journal of Veterinary Sciences* 2022; 53(3): 423-429.
- Then M. Examination on antioxidant activity in the greater celandine (Chelidonium majus L.) extracts by FRAP method. *Acta Biologica Szegediensis*, (2003);47(1-4):115-7.
- Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, et al. Oxidative stress, aging, and diseases. *Clinical interventions in aging*, (2018);13:757.
- Adwas AA, Elsayed A, Azab A, Quwaydir F. Oxidative stress and antioxidant mechanisms in human body. *J Appl Biotechnol Bioeng*, (2019);6(1):43-7.
- Salehi B, Azzini E, Zucca P, Maria Varoni E, V Anil Kumar N, Dini L, et al. Plant-derived bioactives and oxidative stress-related disorders: A key trend towards healthy aging and longevity promotion. *Applied Sciences*, (2020);10(3):947.
- Gorinstein S, Cviková M, Machackova I, Haruenkit R, Park Y-S, Jung S-T, et al. Characterization of antioxidant compounds in Jaffa

- sweeties and white grapefruits. *Food Chemistry*, (2004);84(4):503-10.
27. Ben Salem M, Affes H, Athmouni K, Ksouda K, Dhoubi R, Sahnoun Z, et al. Chemicals compositions, antioxidant and anti-inflammatory activity of *Cynara scolymus* leaves extracts, and analysis of major bioactive polyphenols by HPLC. Evidence-based Complementary and Alternative Medicine, (2017);2017.
 28. Vamanu E, Vamanu A, Nita S, Colceriu S. Antioxidant and antimicrobial activities of ethanol extracts of *Cynara scolymus* (*Cynarae folium*, Asteraceae family). *Tropical Journal of Pharmaceutical Research*, (2011);10(6):777-83.
 29. Sharif KOM, Tufekci EF, Ustaoglu B, Altunoglu YC, Zengin G, Llorent-Martínez E, et al. Anticancer and biological properties of leaf and flower extracts of *Echinacea purpurea* (L.) Moench. *Food Bioscience*, (2021);41:101005.
 30. Wojdyło A, Oszmiański J, Czemerys R. Antioxidant activity and phenolic compounds in 32 selected herbs. *Food chemistry*, (2007);105(3):940-9.
 31. Negahdari, B., Darvishi, M., Saeedi, A.A. Gold nanoparticles and hepatitis B virus. *Artificial Cells, Nanomedicine and Biotechnology*, (2019); 47(1):469-474.
 32. Darvishi M, Ziari K, Mohebi H, Alizadeh K. Association between iron deficiency anemia and *Helicobacter pylori* infection among children under six years in Iran. *Acta Medica Iranica*, (2015); 53(4):220-224.
 33. Jalalmanesh S, Darvishi M, Rahimi M, Akhlaghdoust M. Contamination of senior medical students' cell phones by nosocomial infections: A survey in a university-affiliated hospital in Tehran. *Shiraz E Medical Journal*, (2017); 18(4): 43920.
 34. Darvishi M, Nazer MR, Alipour MR. Investigating the end of patients suffering from diabetic foot hospitalized in Be'sat hospital of IRIAF from 2009 to 2014. *Biomedical Research*, (2017); 28(10): 4360-4633.
 35. Pirhadi M, Shariatifar N, Bahmani M, Manouchehri A. Heavy metals in wheat grain and its impact on human health: A mini-review. *Journal of Chemical Health Risks*, (2021): Apr 6.
 36. Manouchehri A, Ahangar RM, Bigvand P, Nakhaee S, Mehrpour O. Successful treatment of heart failure due to simultaneous poisoning with aluminum phosphide and zinc phosphide: a case report. *Iranian Red Crescent Medical Journal*, (2019): Mar 1;21(3).
 37. Darvishi M, Foroootan M, Nazer MR, Karimi E, Noori M. Nosocomial Infections, Challenges and Threats: A Review Article. *Iranian Journal of Medical Microbiology*, (2020); 14(2):162-181.
 38. Sedighi M, Namdari M, Mahmoudi P, Khani A, Manouchehri A, Anvari M. An Overview of Angiogenesis and Chemical and Physiological Angiogenic Factors: Short Review. *Journal of Chemical Health Risks*, (2022): Jan 15.
 39. Foroootan M, Tabatabaefar M, Mosaffa N, Ashkalak HR, Darvishi M. Investigating esophageal stent-placement outcomes in patients with inoperable non-cervical esophageal cancer. *Journal of Cancer*, (2018); 9(1): 213-218
 40. Darvishi M. Antibiotic resistance pattern of uropathogenic methicillin-resistant staphylococcus aureus isolated from immunosuppressive patients with pyelonephritis. *Journal of Pure and Applied Microbiology*, (2016); 10(4):2663-2667.
 41. Tavakolpour S, Darvishi M, Mirsafaei HS, Ghasemiadl M. Nucleoside/nucleotide analogues in the treatment of chronic hepatitis B infection during pregnancy: a systematic review. *Infectious Diseases*, (2018); 50(2):95-106
 42. Darvishi M, Sadeghi SS. Evaluation of association of *Helicobacter pylori* infection and coronary heart disease (CHD) among CCU patients. *Journal of Pure and Applied Microbiology*, (2016); 10(4):2621-2626.
 43. Manouchehri AA, Pirhadi M, Shokri S, Khaniki GJ. The Possible effects of heavy metals in honey as toxic and carcinogenic substances on human health: A systematic review. *Uludağ Arıcılık Dergisi*, (2021); 21(2):237-46.
 44. Jafari H, Nemati M, Haddad Molayan P, Khaleghi Rostamkoliae L, Hamidi Nejat H. Scolicidal activity of *Mesobuthus eupeus* venom against the protoscolices of *Echinococcus granulosus*. *Archives of Razi Institute*, (2019): 74(2), 183-189.
 45. Salah Najim A, Bahry Al_Sadoon M, Salem Sheet M. Effect of Caraway Seed Extract on the Blood Biochemistry and Antioxidant Capacity among the Hyperoxidative Stress-Induced Rats. *Archives of Razi Institute*, (2022): 77(2), 553-563.
 46. Moharreri M, Vakili R, Oskoueian E, Rajabzadeh G. Evaluation of Microencapsulated Essential Oils in Broilers Challenged with *Salmonella Enteritidis*: A Focus on the Body's Antioxidant Status, Gut Microbiology, and Morphology. *Archives of Razi Institute*, (2022): 77(2), 629-639.
 47. Varadhan S, Venkatchalam R, Perumal S, Ayyamkulamkara S. Evaluation of Oxidative Stress Parameters and Antioxidant Status in Coronary Artery Disease Patients. *Archives of Razi Institute*, (2022): 77(2), 853-859.
 48. Jafari H, Salabi F. Phylogenetic and Morphological Analyses of *Androctonus crassicauda* from Khuzestan Province, Iran (Scorpiones: Buthidae). *Archives of Razi Institute*, (2020): 75(3), 405.
 49. Masoudi R, Dadashpour Davachi N. Effect of Dietary Fish Oil on Semen Quality and Reproductive Performance of Iranian Zandi Rams. *Archives of Razi Institute*, (2021): 76(3), 621-629.
 50. Kamil Kadhim Lawi Z, Ameen Merza F, Rabeea Banoon S, Al-Saady M, Al-Abboodi A. Mechanisms of Antioxidant Actions and their Role in many Human Diseases: A Review. *Journal of Chemical Health Risks*, (2021): 11(1): 45-57.
 51. Durgawale PP, Patil MN, Joshi SA, Korabu KS, Datkhile KD. Studies on phytoconstituents, in vitro antioxidant, antibacterial, antiparasitic, antimicrobial, and anticancer potential of medicinal plant *Lasiosiphon eriocephalus decne* (Family: Thymelaeaceae). *Journal of Natural Science, Biology and Medicine*, (2019): 1;10(1):38.
 52. You SH, Yoon MY, Moon JS. Antioxidant and Anti-inflammatory Activity Study of Fulvic Acid. *Journal of Natural Science, Biology and Medicine*, (2021): 12(3): 1.
 53. Datkhile KD, Patil SR, Patil MN, Durgawale PP, Jagdale NJ, Deshmukh V. N. Studies on phytoconstituents, In vitro antioxidant, antibacterial, and cytotoxic potential of *Argemone mexicana* Linn. (Family: Papaveraceae). *Journal of Natural Science, Biology and Medicine*, (2020): 11(2), 198.
 54. Ramesh C, Vinithkumar NV, Kirubakaran R. Marine pigmented bacteria: A prospective source of antibacterial compounds. *Journal of Natural Science, Biology and Medicine*, (2019): 10(2), 104.
 55. Halim S, Jasmi NA, Ridzuan PM, Anna D, Abdullah S, Sina T. Novel potential *Centella asiatica* extract in ameliorating neurotoxicity induced oxidative stress in chronic morphine dependant rat model. *International Journal of Medical Toxicology & Legal Medicine*, (2020): 23(3and4), 79-83.
 56. Othman Z, Khalep HRH, Abidin AZ, Hassan H, Fattapur S. The Anti-Angiogenic Properties of *Morinda citrifolia* L (Mengkudu) Leaves using chicken chorioallantoic membrane (CAM) assay. *Pharmacognosy Journal*, (2019) 11(1): 1.
 57. Fattapur S, Kiran Chanabasappa Nilugal T. Theva Darshan, May Florence Dela Cruz Bacayo, Fadli Asmani, Ibrahim Abdullah, Eddy Yusuf, and Prakash Goudanavar. "Toxicological and pharmacological activity of ethanolic extracts of *Catharanthus roseus* in experimental animals." *International Journal of Medical Toxicology & Legal Medicine*, (2018): 21, no. 3and4: 141-144.
 58. Solati K, Karimi M, Rafieian-Kopaei M, Abbasi N, Abbaszadeh S, Bahmani M. Phytotherapy for wound healing: The most important herbal plants in wound healing based on Iranian ethnobotanical documents. *Mini-Reviews in Medicinal Chemistry*, (2020); 21(4): 500-519.
 59. Bahmani M., Jalilian A, Salimikia I, Shahsavari S, Abbasi N. Phytochemical screening of two Iran native plants *Ziziphus nummularia* (Burm.f.) Wight & Arn. and *Ziziphus spina-christi* (Mill.) Georgi using HS-SPME and GC-MS spectroscopy. *Plant Science Today*, (2020); 7(2): 275-280.
 60. Abbasi N, Khalighi Z, Eftekhari Z, Bahmani M. Extraction and phytoanalysis of chemical compounds of *Eucalyptus globulus* leaf native to Dehloran, Ilam province, Iran by HS-SPME and GC-MS. *Advances in Animal and Veterinary Sciences*, (2020); 8(6): 647-652.

61. Aidy A, Karimi E, Ghaneialvar H, Mohammadpour S, Abbasi N. Protective effect of Nectaroscordum tripedale extract and its bioactive component tetramethylpyrazine against acetaminophen-induced hepatotoxicity in rats. *Advances in Traditional Medicine*, (2020); 20(3): 471-477.
62. Abbasi N, Khosravi A, Aidy A, Shafiei M. Biphasic response to luteolin in MG-63 osteoblast-like cells under high glucose-induced oxidative stress. *Iranian Journal of Medical Sciences*, (2016); 41(2): 118-125.
63. Uddin MK, Juraimi AS, Anwar F, Hossain MA, Alam MA. Effect of salinity on proximate mineral composition of purslane (*Portulaca oleracea* L.). *Australian journal of crop Science*, (2012): 6(12):1732-6.
64. Alam M, Juraimi AS, Rafii M, Abdul Hamid A, Aslani F, Hasan M, et al. Evaluation of antioxidant compounds, antioxidant activities, and mineral composition of 13 collected purslane (*Portulaca oleracea* L.) accessions. *BioMed Research International*, (2014); Article ID 296063.



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